

## Article

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## **Title Page**

# **Hyperglycaemia-Induced Cardiac Contractile Dysfunction in the Diabetic Heart**

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## **Abstract**

The development of a diabetic cardiomyopathy is a multifactorial process, and evidence is accumulating that defects in intracellular free calcium concentration  $[Ca^{2+}]_i$  or its homeostasis are related to impaired mechanical performance of the diabetic heart leading to a reduction in contractile dysfunction. Defects in ryanodine receptor, reduced activity of the sarcoplasmic reticulum calcium pump (SERCA) and, along with reduced activity of the sodium-calcium exchanger (NCX) and alterations in myofilament, collectively cause a calcium imbalance within the diabetic cardiomyocytes. This in turn is characterized by cytosolic calcium overloading or elevated diastolic calcium leading to heart failure. Numerous studies have been performed to identify the cellular, subcellular and molecular derangements in diabetes-induced cardiomyopathy (DCM), but the precise mechanism(s) is still unknown. This review focuses on the mechanism behind DCM, the onset of contractile dysfunction and the associated changes with special emphasis on hyperglycaemia, mitochondrial dysfunction in the diabetic heart. Further, management strategies, including treatment and emerging therapeutic modalities are discussed.

**Key words:** Contractile dysfunction, sarcoplasmic reticulum calcium ATPase (SERCA), hyperglycaemia, diabetic cardiomyopathy, calcium transients  $[Ca^{2+}]_i$

## **Diabetes mellitus (DM)**

Diabetes mellitus (DM) is a heterogeneous metabolic disorder characterised by chronic hyperglycaemia (HG) that results in dysfunction in the cell's ability to transport and utilize glucose [1]. Type 1 DM (T1DM), is caused by T lymphocyte-mediated autoimmune destruction of the pancreatic  $\beta$ -cells, resulting in insufficient insulin production and subsequent decrease in glucose utilization [2]. Type 2 DM (T2DM) results from an insulin resistance that instigates hypertrophy of the  $\beta$ -cell to compensate, and resulting in hyperinsulinemia leading to eventual insulin resistance [3, 4]. Advanced failure of the  $\beta$ -cells in T2DM decreases the amount of insulin produced resulting in pathophysiological changes that produce elevated blood glucose levels (hyperglycaemia), impaired cellular glycolysis and pyruvate oxidation [5]. In chronic hyperglycaemia, comorbidities include some long-term complications as peripheral vascular disease, retinopathy, kidney failure, neuropathy and eventually, cardiac dysfunction/failure or cardiomyopathy [6].

Currently, more than 422 million people worldwide – almost one in every 11, suffer from DM and this number is likely to increase to 650 million within the next 15-20 years [7], making it one of the five leading causes of death in developed and developing countries. Between 1980 and 2014 the number of adults with diabetes quadrupled from 108 million to 422 million. The prevalence of DM was higher in the developed countries but now, the major increase in people with diabetes occurs in the developing countries such as China, India, and those in Middle East and Africa.

Type 2 DM (T2DM) is specifically epidemic due to the rising rates of obesity throughout the world. Over one billion people worldwide are overweight (BMI >25 and <29.9) or obese (BMI >30) in most cases and they are also pre-diabetics [8]. With such high rates of morbidity and mortality, DM represents a major medical, sociological and economic challenge and burden to the national health services globally.

Cardiovascular diseases (CVDs) are the leading cause of death throughout the world and on the basis of 2011 death rate data, mortality owing to cardiovascular disease accounted a astounding 2150 people dying daily with an average of 1 death every 40 seconds [9]. Interesting, CVD- related mortality with DM have been reported to be about 65% [10] and 68% of adults with DM older than 65 years die of some form of heart disease. The Framingham Heart Study demonstrated that the frequency of heart failure is five times greater in diabetic women and two times greater in diabetic men compared with age-

matched control subjects [11,12].

Diabetic heart disease is now recognized as an important and growing public health risk which affects the heart in three ways: cardiac autonomic neuropathy (CAN), coronary artery disease (CAD) due to accelerated atherosclerosis, and diabetic cardiomyopathy (DCM) [10].

DCM is a heart failure syndrome found in diabetic patients that is characterized by impaired myocardial relaxation dynamics or diastolic dysfunction, and structural abnormalities leading to left ventricular hypertrophy (LVH) or a combination of these [13]. Presently, DCM is defined as myocardial dysfunction (MD) in patients with DM in the absence of hypertension and structural heart diseases such as valvular heart disease or CAD [14].

Other features of DCM include: interstitial fibrosis, myocyte hypertrophy [15], lipid accumulation in cardiomyocytes and fetal gene reactivation [16]. Furthermore, DCM is also accompanied by comorbidities such as obesity, smoking, hypertension and others and these complications often precede the development of systolic dysfunction, CAD and heart failure [17].

With DM being a well-known risk factor for the development of heart failure, which leads to a poor quality of life in affected individuals, the situation now becomes complicated in treatment of DM resulting in alteration of the pharmacokinetics of anti-diabetic medications. Therefore, in this review we focus on the mechanism behind DCM, the onset of contractile dysfunction and the associated changes with special emphasis on hyperglycaemia, mitochondrial dysfunction in the diabetic heart and management strategies, including treatment and emerging therapeutic directions.

## **Mechanism of DCM**

Several factors are involved in the development of diabetic cardiomyopathy, including metabolic, biochemical, and ultrastructural changes within the cardiac myocyte [18,19]. In particular, because of the primary role of mitochondria in ATP production, weakening of mitochondrial respiratory function is a key contributing factor to decreased contractile function of the diabetic heart. Numerous mechanisms have been proposed to contribute to this clinical situation, including oxidative stress, microvascular abnormalities, and decreased sarcoplasmic reticular calcium uptake [20]. Other proposed mechanisms include: subcellular component abnormalities, metabolic disturbances, cardiac autonomic dysfunction, alterations in the renin-angiotensin-aldosterone system (RAAS), and maladaptive immune

responses [21]. However, the molecular mechanisms that cause this cardiac dysfunction are still largely undefined. An established hypothesis is that hyperglycemia plays a critical role in the development of DCM, though multiple complex mechanisms and the interplay of many metabolic and molecular events within the myocardium and plasma contribute to its pathogenesis. The principal metabolic abnormalities in DM are hyperglycemia and hyperlipidemia, all of which stimulate the production of reactive oxygen species (ROS) or the nitrogen species that cause most diabetic complications, including DCM and diabetic nephropathy [22]. These abnormalities further induce alterations in downstream transcription factors, which result in changes in gene expression, myocardial substrate utilization, myocyte growth, endothelial function and myocardial compliance.

The development of DCM is a multifactorial process, and accumulating evidence has revealed that defects in  $[Ca^{2+}]_i$  homeostasis are related to impaired mechanical performance of the diabetic heart leading to the prevalence of contractile dysfunction [23]. It has been suggested that diastolic dysfunction may be due to cardiomyocyte hypertrophy and myocardial fibrosis. In turn, at cellular level, these are associated with defects in calcium transport and mitochondrial calcium uptake due to mitochondrial dysfunction and reduced activity of SERCA pump which are all responsible for calcium sequestration during cardiomyocyte diastolic relaxation [24-26]. Also, there is much evidence that reduced activity of NCX, myocardial contractile protein collagen formation and fatty acid metabolism are also associated with diastolic dysfunction. *In vivo* functional changes include decreased ventricular filling, decreased ventricular ejection fraction, decreased fractional shortening, increased ventricular wall stiffness and increased pre-ejection time [17].

As stated in the aforementioned section, hyperglycaemia stimulates an increase in reactive oxygen species (ROS) and reactive carbonyl species (RCS) production, because of increased input of reducing equivalents into the mitochondrial electron transport chain [27-28]. Hyperglycaemia-induced cell damage is a consequence of increased flux through metabolic pathways (polyol pathway flux, advanced glycation- end product formation, activation of protein kinase C (PKC) isoforms and increased hexosamine pathway flux) [22,29]. Mitochondrial function is highly impaired by hyperglycaemic conditions in part because of decreased mitochondrial transcription factor A (TFAM) activity and/or expression [30-31]. These topics will be discussed more in-depth in subsequent sections.

## Glucose metabolism regulation by insulin and fatty acids

It was reported over 40 years ago [32] that glycolytic metabolism is increased in cardiac hypertrophy and congestive heart failure and it is now well-known that cardiac glucose metabolism declines in diabetes [33] due to a decline in insulin, insulin resistance or increased availability of fatty acids. Healthy hearts derive most of their energy from free fatty acids and only a small proportion from circulating glucose while; in contrast, diabetic hearts use more fat and less glucose than normal hearts.

The diabetic heart is characterized by distinctive metabolic events including elevations in fatty acid uptake and oxidation combined with a decrease in glucose uptake and oxidation. A major factor in the elevated uptake and oxidation of fatty acids in cardiomyocytes has been linked to increased release of fatty acids by the adipocyte and liver resulting in elevated circulating fatty acids and triglycerides. [34]. In the heart, utilization of glucose stimulated by insulin, is inhibited by fatty acid metabolism *via* transcriptional regulation of limiting enzymes. The generation of enzymes of fatty acid oxidation entails the transcriptional regulators PPAR $\alpha$  and/or PPAR $\beta$ , which regulate gene expression of these enzymes and are elevated and activated in diabetic hearts [25]. It has been suggested that the presence of glucose reduces fatty acid metabolism, probably by increasing intracellular levels of malonyl CoA [36], while increased fat dependence also appears to play a role in the function of decreased glucose metabolism. The initial step in cardiomyocyte glucose usage is uptake, which is significantly regulated by insulin in the heart. In DM there is also a persistent reduction in cardiac glycolytic capacity [37] and glucose oxidation is further reduced by a decline in pyruvate dehydrogenase activity [37,38].

The concept of limiting enzymes and limiting steps in glucose metabolism has been reported to contribute to development of DCM [27] where limiting steps have become potential sites of pathology. The concept of one such limiting step was explained by Randle et al. [32] in the 1960's and more recently by An and Rodrigues [5] relating to glucose uptake *via* the glucose transporters GLUT1 and GLUT4 in myocardial glucose metabolism. These authors underlined that enabling of glucose transport by insulin involves both the translocation of GLUT4 to the cell membrane and the upregulation of the transporter, an effect that is impaired in diabetes. It was reported that despite the significant reduction in glucose uptake, the size of the intracellular glucose pool is elevated in the type 1 diabetic heart.

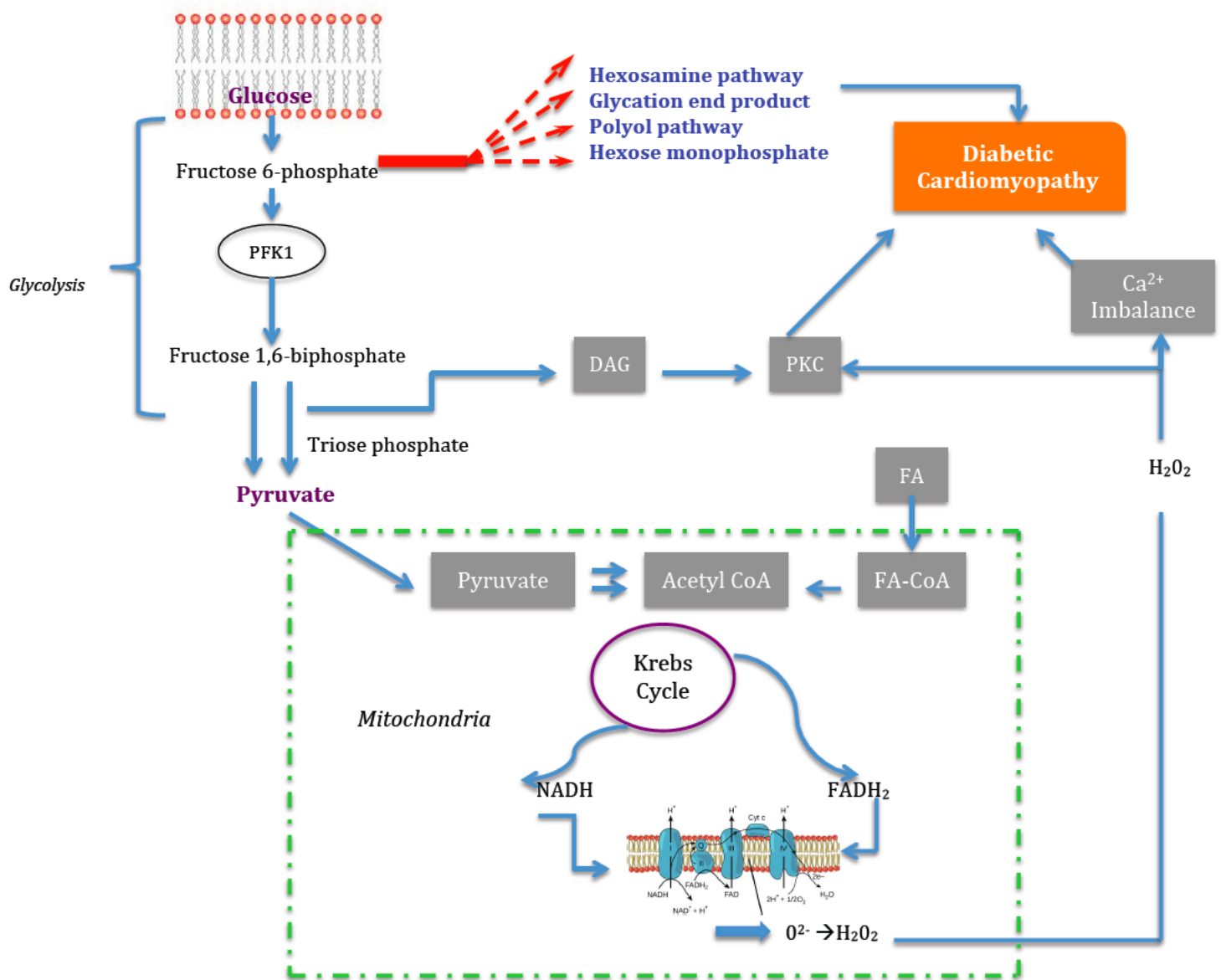
[31]. Data collected revealed that impaired GLUT4 activity in the diabetic heart does not limit glycolytic flux and would imply that key bottlenecks in glycolysis develop downstream from the glucose transporters in the diabetic heart.

Glycolysis is the metabolic pathway that involves a sequence of ten enzyme-catalysed reactions, which converts glucose into pyruvate. One such important limiting enzyme in the diabetic heart is phosphofructokinase (PFK), which catalyses the conversion of fructose-6-phosphate to fructose 1,6 bisphosphate. Enhanced rates of fatty acid  $\beta$  -oxidation lead to elevations in cardiac levels of both acetyl CoA and citrate, and it has been shown previously that citrate is a potent inhibitor of PFK [40]. Thus, this enzyme is a key target of fatty acid-mediated regulation of glycolysis. Owing to the fact that diabetes is associated with enhanced rates of fatty acid  $\beta$  - oxidation and elevations in citrate levels, it has been proposed that PFK activity is diminished in the diabetic heart [40]. Further research also implicates this enzyme as a major sensor of the high-energy phosphate content of the heart [41]. Elevations in AMP kinase activity as well as low ATP/ADP ratio has shown to stimulate the activity of this allosteric enzyme.

Other researchers [42] have proposed data supporting a role for PFK in diabetes-mediated glycolytic impairment that correlate with elevations in glucose-6-phosphate levels and reductions in fructose 1,6 bisphosphate content. Based upon these findings, a bottleneck consequently develops in the glycolytic pathway of the diabetic heart, resulting in an increase in myocardial levels of glucose-6-phosphate and fructose-6- phosphate. These events have important pathological consequences, as they serve as substrates for four pathological pathways involved in the development of DCM (see Figure 1).

Other limiting steps/enzymes in regulation of glucose metabolism in the heart include: (I) the three- enzyme pyruvate dehydrogenase complex (PDH) and (II) Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) – especially during myocardial ischemia.





**Figure 1:** Flow diagram showing metabolic mechanisms of diabetic cardiomyopathy and how ROS from the mitochondria can leave the mitochondria as H<sub>2</sub>O<sub>2</sub> and activate PKC in the cytosol. PFK- phosphofructokinase; DAG – diacylglycerol; PKC – protein kinase C; FA – fatty acid; NADH – nicotinamide adenine dinucleotide (reduced form); FADH – flavin adenine dinucleotide (reduced form); diagram drawn by hand.

### Mitochondrial dysfunction and changes in PKC activities in the diabetic heart

Mitochondrial acetyl Co-A is a crucial factor of the citric acid cycle. In mammalian cells, mitochondrial acetyl-CoA is produced from pyruvate or by the  $\beta$ -oxidation of fatty acids, and

then feeds into the citric acid cycle. Since the 1960's researchers [43] have highlighted the important role of acetyl-CoA in diabetic heart. Bowman [43] concluded that elevated levels of citric acid cycle intermediates in hearts of diabetic rats and in normal hearts perfused with fatty acids and ketone bodies are due largely to increased availability of acetyl-CoA.

Another factor that plays an important determinant role in the citric acid cycle flux is slowing of the respiratory chain flux. Several authors have suggested [44-47] that this event would contribute to the inhibition of  $\alpha$ -ketoglutarate dehydrogenase and elevate the NADH/NAD<sup>+</sup> ratio, which in turn, would result in diminished citric acid flux.

Complexes in the respiratory chain are determined by the transcriptional activity of mitochondrial transcription factor A (TFAM), which is essential for mitochondrial DNA transcription and replication. It has been proposed [48] that TFAM transcriptional activity is decreased in diabetic cardiomyopathy and TFAM activity may be responsible for some of the alterations caused by hyperglycaemia. These researchers [48] investigated the effect of TFAM overexpression on hyperglycaemia-induced cytosolic calcium handling and mitochondrial abnormalities. They discovered that overexpression of TFAM dramatically affects the function of neonatal rat cardiomyocytes incubated in medium containing 30 mM glucose. ATP content was reduced by 30% and mitochondrial calcium decreased by 40% after high glucose. Calcium transients were prolonged by 70% after high glucose, which was associated with diminished sarco(endo)plasmic reticulum Ca<sup>2+</sup>-ATPase 2a and cytochrome-c oxidase subunit 1 expression. In summary, it was demonstrated that: (I) hyperglycaemia increases the levels of O-GlcNAcylated TFAM, which diminishes the activity of TFAM and reduces the activity of oxidative phosphorylation and (II) TFAM overexpression protected cell function against the damage induced by high glucose in cardiomyocytes.

An additional consequence of weakened respiratory chain flux is the generation of reactive oxygen species (ROS) – produced as electrons are diverted from the respiratory chain to the acceptor oxygen.

**Figure 1** shows how ROS (O<sup>2•</sup>) generated from the mitochondria can leave the organelles as H<sub>2</sub>O<sub>2</sub> and activate PKC in the cytosol. The outcome of PKC activation by ROS, in turn, triggers activation of NADPH oxidase and results in considerably increased cytosolic ROS content. Consequently, diabetes-mediated mitochondrial dysfunction contributes to the development of DCM by altering ATP generation and Ca<sup>2+</sup> movement.

Protein kinase C (PKC) has been shown to inhibit sarcoplasmic reticular  $\text{Ca}^{2+}$  pump and myofibrillar ATPase [49,50] activities, and several investigators have suggested that subcellular changes in the diabetic heart may be due to alterations in the PKC activity and/or PKC-mediated signal transduction mechanism. Some examples include (I) increased phosphorylation of troponin-I in the diabetic heart has been considered to be due to the activation of PKC [51,52] and (II) diabetes is related with translocation of the  $\epsilon$ -isoform of PKC from cytosolic to particulate fraction of cardiomyocytes. This change is prevented by the blockade of angiotensin II receptors, which are known to activate PKC [51] and it has also been established that concentration of diacylglycerol, a known activator of PKC [53] was increased in the diabetic heart.

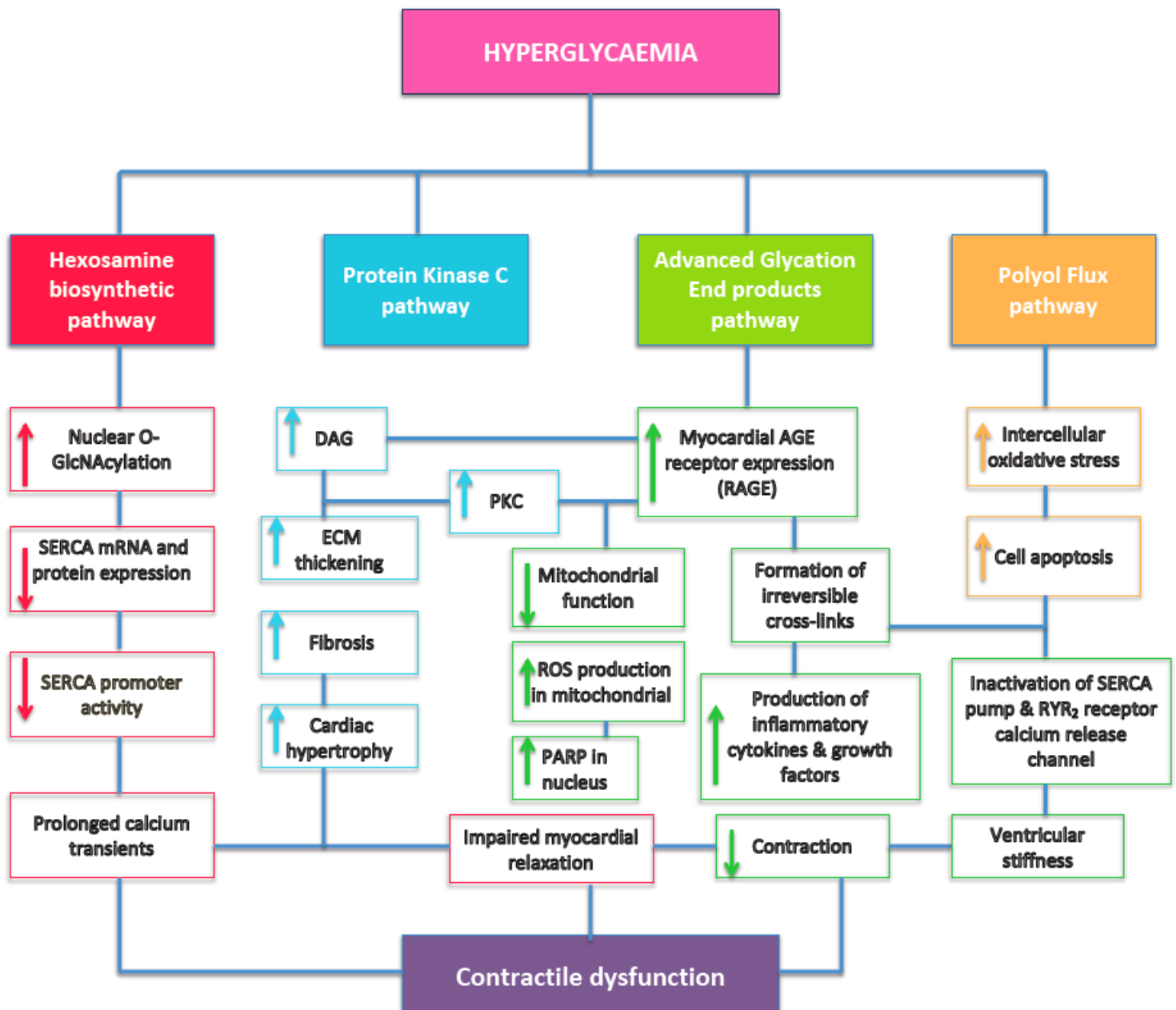
One important study undertaken by Liu et al. [54] examined PKC activities in the homogenate, cytosolic, and particulate fractions from diabetic hearts. They found  $\text{Ca}^{2+}$ -dependent PKC activity was increased by 43 and 51% in the homogenate fraction and 31% and 70% in the cytosolic fraction from the 4- and 8-weeks diabetic hearts while  $\text{Ca}^{2+}$ -independent PKC activity was increased by 24% and 32% in the homogenate fraction and 52% and 89% in the cytosolic fraction respectively, both cases in comparison with control values. They also examined changes in the contents of different PKC isozymes in cardiac homogenate, cytosolic, and particulate fractions in diabetes induced by streptozotocin (STZ) in rats. The results showed relative protein contents of PKC- $\alpha$ - $\beta$ - $\epsilon$  and  $\zeta$ , isozymes were increased by 43%, 31%, 48%, and 38%, respectively, in the homogenate fraction and by 126%, 119%, 148%, and 129%, respectively, in the cytosolic fraction of the 8-weeks diabetic heart. These results provided reliable evidence that the increased myocardial PKC activity and increased protein contents of the cytosolic PKC isozymes are associated with subcellular alterations and cardiac dysfunction in the diabetic heart.

For more in-depth review and explanations on regulation of myocardial glucose, fatty acid metabolism diseases and dysfunctions, several excellent review articles have been published in the area of diabetes induced cardiac dysfunction [55-59].

## **Role of HG and its biochemical pathways in the development of cardiac dysfunction**

Various mechanisms of HG are responsible for the generation of diabetes-induced heart disease, including metabolic abnormalities such as cellular calcium overload and altered

calcium metabolism in cardiomyocytes leading to contractile dysfunction [60-63]. One major contributor to HG-induced diabetic abnormalities is increased oxidative stress along with depleted antioxidant defences and raised levels of reactive oxygen species (ROS). Persistent hyperglycaemia results in increased glucose metabolism, which increases oxidative stress *via* the development of ROS from the mitochondria. ROS can cause damage to the mitochondria together with poly (ADP-ribose) polymerase-1 (PARP) activation leading to the inhibition of the cytosolic enzyme glyceraldehyde-3-phosphate dehydrogenase (GAPDH). This inhibition initiates a series of cellular processes, by activating pathways that lead to HG-associated cellular/tissue damage [64]. Inhibition of GAPDH diverts glucose from glycolytic pathways into alternative biochemical pathways, including advanced glycation end product (AGE) formation, hexosamine, polyol pathway, and protein kinase C (PKC) activation [65]. Increased formation of AGEs is involved in forming irreversible cross-links with macromolecules such as collagen leading to myocardial fibrosis, inactivation of SERCA2a and RyR2 calcium release channel, together with impaired cardiac contractility, relaxation and ventricular stiffness [66-69]. Increased polyol flux is associated with reduced levels of intracellular glutathione and an increase in cardiac cell apoptosis [70]. Furthermore, inhibition of this pathway has been claimed to provide protection for the heart from ischaemic injury [71]. The hexosamine biosynthetic pathway is known for reducing SERCA2a mRNA and protein expression, along with reduced SERCA2a promoter activity via increased nuclear O-GlcNAcylation. This results in prolonged calcium transients and impaired myocardial relaxation [72-73]. Finally, the activation of protein kinase C (PKC) *via* de novo synthesis of the lipid second messenger diacylglycerol (DAG) leads to vascular alterations at pathological, cellular and functional levels which include basement membrane thickening, extracellular matrix expansion, vascular permeability, enzymatic alterations such as Na<sup>+</sup>- K<sup>+</sup>-ATPase, and MAP kinase, multifocal fibrosis, myocyte necrosis, decreased left ventricular performance and left ventricular hypertrophy [74]. Impaired calcium handling and cellular efflux may further contribute to impaired relaxation, or diastolic dysfunction. Overall, HG *via* multiple biochemical pathways results in myocardial, cellular and functional changes, all which contribute to the development of a cardiomyopathy leading to HF. Figure 2 illustrates these pathways as a flow chart.



**Figure 2:** A schematic flow chart representation highlighting the role of hyperglycaemia and its biochemical pathways including hexosamine biosynthetic pathway, Protein Kinase C (PKC) pathway, advanced glycation end-products pathway and the Polyol flux pathway in the development of cardiac dysfunction. {DAG; diacylglycerol, ECM; Extracellular Matrix, PARP; poly (ADP-ribose) polymerase-1, SERCA; sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase}. See text for discussion; Diagram drawn by hand.

## Regulation of calcium in normal cardiac muscle

The major function of the heart is to pump blood efficiently by virtue of an orchestrated contraction–relaxation cycle of the working cardiomyocytes. Contractility of these

cardiomyocytes is regulated by a spatially defined programme of ion channels and exchangers that precisely control  $\text{Ca}^{2+}$  entry in and out of the cell and the SR. Regulation of contractility and hence, the control of  $\text{Ca}^{2+}$  release is principally achieved *via* the electrical activity of the sarcolemma. Depolarization of the cardiac cell membrane, during a normal action potential is sustained in the plateau phase by the activation of voltage-gated L-type  $\text{Ca}^{2+}$  channels (ICa,L) [75]. It is this small influx of  $\text{Ca}^{2+}$  *via* these channels that triggers a much larger release of  $\text{Ca}^{2+}$  from the SR by a process called calcium–induced calcium release (CICR). Upon activation of the SR, there is a transient rise in cytoplasmic  $\text{Ca}^{2+}$  concentration  $[\text{Ca}^{2+}]_i$ . This phenomena is commonly referred to as the calcium transient  $[\text{Ca}^{2+}]_i$  and this CICR process is generally accepted as the major mechanism of  $\text{Ca}^{2+}$  release from the SR.  $\text{Ca}^{2+}$  release from the SR is mediated by intracellular calcium receptors commonly known as ryanodine receptors (RyR), with type 2 RyRs being the most abundant intracellular  $\text{Ca}^{2+}$  channels in cardiomyocytes [75,76]. Contraction is initiated when free  $\text{Ca}^{2+}$  causes the interaction of the myofilaments *via* troponin C and the thick and thin filaments, namely actin and myosin leading to cell shortening. Exclusion of  $\text{Ca}^{2+}$  from the cytosol is achieved mainly by several mechanisms, including SR uptake via the SR  $\text{Ca}^{2+}$  transporter (SERCA), removal through the sarcolemma *via* the NCX and to a small extent by the  $\text{Ca}^{2+}$ -ATPase pump [77-78]. These changes result in both cyclic increases and decreases in  $\text{Ca}^{2+}$  and in contractility of the individual myocytes.

In diabetic cardiomyopathy, it is well known that  $\text{Ca}^{2+}$  homeostasis is deranged leading to elevated diastolic  $\text{Ca}^{2+}$ . This is due to a failure of SERCA to pump back  $\text{Ca}^{2+}$  into the SR, increased asynchronous SR  $\text{Ca}^{2+}$  leak *via* RyRs and dysfunction of the NCX. These three  $\text{Ca}^{2+}$  transport proteins participate during calcium transient  $[\text{Ca}^{2+}]_i$  decline leading to a slow steady-state in resting  $\text{Ca}^{2+}$  [77]. In the new balanced state, a generously proportioned fraction of activating  $\text{Ca}^{2+}$  also enters the cell at each beat via the L-type  $\text{Ca}^{2+}$  channel (e.g. smaller  $\text{Ca}^{2+}$  release causes less  $\text{Ca}^{2+}$  inactivation). Consequently, these 4 major transporting proteins have been identified and they seem to contribute to the disturbed diastolic  $\text{Ca}^{2+}$  accumulation observed in the failing heart. Firstly, increased  $\text{Ca}^{2+}$  leak through RyR, secondly reduced SERCA activity, decreased trans-sarcolemal elimination of  $\text{Ca}^{2+}$  by the NCX and lastly the L-type  $\text{Ca}^{2+}$  channel [79-80]. These issues are further examined in the following text.

### **Ryanodine receptor dysfunction**

A major feature in diabetic cardiomyopathy is the increased  $\text{Ca}^{2+}$  leak from the SR due to enhanced RyR open probability. Because leak, as measured by  $\text{Ca}^{2+}$  sparks, is increased with

higher SR  $\text{Ca}^{2+}$  load, any intervention to increase SR  $\text{Ca}^{2+}$  load without reducing the leak decreases efficiency of ECC process with increased energy consumption with arrhythmias as potential side effects [79,81].

RyRs were first observed in the 1970's as 'foot' structures in electron micrograph images of striated muscle filling the gaps that are found at specific junctions between the sarcolemma and the SR membrane [82-83]. Lai and colleagues identified the RyR as an integral SR membrane protein with a role in  $\text{Ca}^{2+}$  release [84]. The complementary DNA encoding three distinct RyR channels was cloned and the corresponding gene sequences obtained for three isoforms; RyR1 [85-86], RyR2 [87-88], and RyR3 [89]. It was not until the 1990s that the central role of RyRs through numerous biochemical, physiological, molecular and pharmacological studies and the physiological characteristics of excitation–contraction coupling (ECC) were recognized [89-91]. During the past decade numerous discoveries have been made of RyR2 gene mutations, which underlie the arrhythmogenesis leading to sudden cardiac death, which has added a new focus to the role of RyR2 dysfunction in cardiac disease [79-80, 92-93].

### **Dysregulation of sarcoplasmic reticulum and $\text{Na}^+/\text{H}^+$ , $\text{Na}^+/\text{K}^+$ & $\text{Na}^+/\text{Ca}^{2+}$ exchange**

The sarcoplasmic reticulum (SR) constitutes the main intracellular calcium store in striated muscle and it plays an important physiological role in the regulation of excitation-contraction-coupling (ECC) and of intracellular calcium concentrations during contraction and relaxation. Intracellular  $\text{pH}$  and  $[\text{Na}^+]$  in the heart are regulated by the sarcolemmal membrane  $\text{Na}^+/\text{H}^+$  exchange pathway. DCM is characterized by reduced cardiac contractility due to direct changes in myocardium function independent of vascular disease. It is now becoming clear that cardiac dysfunction in chronic diabetes is intimately involved with  $\text{Ca}^{2+}$ -handling abnormalities in the diabetic heart. These abnormalities occur mainly due to defects in sarcolemmal  $\text{Na}^+/\text{K}^+$  ATPase,  $\text{Na}^+/\text{Ca}^{2+}$  exchange,  $\text{Na}^+/\text{H}^+$  exchange,  $\text{Ca}^{2+}$ -channels and  $\text{Ca}^{2+}$ -pump activities as well as changes in sarcoplasmic reticular  $\text{Ca}^{2+}$ -uptake and  $\text{Ca}^{2+}$ -release processes; these alterations may lead to the occurrence of intracellular  $\text{Ca}^{2+}$  overload.

This section investigates the alterations of cardiac sarcoplasmic reticulum  $\text{Ca}^{2+}$ -ATPase activity and cardiac function.

Early innovative work performed by Ganguly et al. [94] investigated the defective sarcoplasmic reticular calcium transport in diabetic cardiomyopathy. The results of this study provided some evidence that the depression in cardiac sarcoplasmic reticular calcium accumulation during diabetes is a consequence of insulin deficiency and associated chronic metabolic changes, but the hypothyroid condition that accompanies experimental diabetes does not appear to play any role in this defect. In follow-up research [95], experiments were designed to monitor rats that were injected with streptozotocin (65 mg/kg), killed 8–10 weeks later, and sarcolemmal membrane vesicles isolated from pooled ventricles. The results from these studies showed significant depressions in  $\text{Na}^+/\text{K}^+$ -adenosine triphosphatase (ATPase) activity and  $\text{Na}^+/\text{Ca}^{2+}$  exchanges were observed in the diabetic preparations in comparison to control. Further, a striking depression in cardiac sarcolemmal  $\text{Na}^+/\text{H}^+$  exchange was observed in the diabetic animals in comparison to control.] Other studies, [96] focused on sarcolemmal  $\text{Ca}^{2+}$  transport in streptozotocin-induced diabetic cardiomyopathy in rats, provided results that indicated a depression in the ability of the cell to remove  $\text{Ca}^{2+}$  through  $\text{Na}^+/\text{Ca}^{2+}$  exchange and  $\text{Ca}^{2+}$ -pump mechanisms in sarcolemma. It was further concluded that these defects might contribute to the occurrence of intracellular  $\text{Ca}^{2+}$  overload and diabetic cardiomyopathy. Lun et al. [97] examined alterations in  $\text{Ca}^{2+}$ -channels during the development of diabetic cardiomyopathy with specific binding of 3H-nitrendipine studied at different concentrations. The outcome of their studies showed significant decrease in both dissociation constant and maximal number of 3H-nitrendipine binding observed after 3 and 8 weeks. Also, it was revealed that treatment of diabetic animals with insulin prevented the occurrence of these changes in the myocardium. In summary, a relationship was seen between number of 3H-nitrendipine binding sites and increased affinity of  $\text{Ca}^{2+}$  channels with the former partly explaining the depressed cardiac contractile force development in chronic diabetes, and latter, partly explaining the increased sensitivity of diabetic heart to  $\text{Ca}^{2+}$ .

Further work by Dhalla et al. [98] highlighted the fact that while reductions in sarcoplasmic reticular  $\text{Ca}^{2+}$  pump and  $\text{Ca}^{2+}$  release channel function are associated with cardiac dysfunction, alterations in sarcolemmal  $\text{Na}^+/\text{Ca}^{2+}$  exchanger and  $\text{Na}^+/\text{K}^+$  ATPase activities contribute to intracellular  $\text{Ca}^{2+}$  overload at late stages of diabetes. The constant buildup of  $\text{Ca}^{2+}$  in mitochondria produces  $\text{Ca}^{2+}$  overload in these organelles, and this change induces impairment of energy production, depletion of energy stores, as well as further promotion of oxidative stress in chronic diabetes. These observations, coupled with generation of



oxyradicals results in the opening of mitochondrial pores, leakage of toxic proteins and myocardial cell damage in diabetes, supports the view that alterations in sarcoplasmic reticular and mitochondrial functions produce intracellular  $\text{Ca}^{2+}$  overload and therefore, play an critical role in the development of cardiac dysfunction in DCM.

Numerous researchers conducted other studies on the sarcoplasmic reticulum transport mechanism. Some of these are explained in the following text.

In cardiac muscle, the SR has been shown to be the most active subcellular organelle implicated with the sequestration of activator calcium. SERCA 2a ATPase, together with sarcolemma NCX are responsible for lowering  $[\text{Ca}^{2+}]_i$ , leading to the relaxation of muscle. Experiments have shown that calcium binding and/or uptake by cardiac SR is altered in a variety of physiological and pathological states [99-101].

Studies involving molecular cloning have identified a family of SERCA pumps encoded by the three homologous genes, SERCA1, SERCA2 and SERCA3. Of the 3 genes. The gene of interest here is the SERCA2, which is alternately spliced and encodes SERCA2a and SERCA2b isoforms [102]. SERCA2b isoform is expressed ubiquitously and is associated with inositol triphosphate (IP3) gated  $\text{Ca}^{2+}$  stores, whereas SERCA2a is the primary isoform expressed in the heart [103]. SERCA-2a ATPase is the major protein pump involved in the process of calcium reuptake into the sarcoplasmic reticulum [103]. Previous studies employing rats, mouse and rabbits have demonstrated that the expression of SERCA pump gradually increases during development [104-105]. Several other studies have shown that DCM is associated with decreased contractility and impaired relaxation [61]. As explained, the expression levels of SERCA pump protein appear to be a critical determinant of cardiac contractility. SERCA2a mRNA expression and protein level and activity have been demonstrated to be down-regulated in streptozotocin-induced (STZ) type 1 DM and the alteration of SERCA expression is accompanied by functional changes [105-106]. Studies from many laboratories have demonstrated that the expression level of SERCA is significantly decreased in pressure overload (Po) induced hypertrophy and HF [105-107]. Within these studies, the main finding has been decreased SR  $\text{Ca}^{2+}$  transport and function. In addition to animal studies on cardiac diseases, there is considerable evidence for alterations in SR  $\text{Ca}^{2+}$  transport and function. In end stage human HF, intracellular  $\text{Ca}^{2+}$  measurements using Fura-2 have shown markedly prolonged  $\text{Ca}^{2+}$  transients  $[\text{Ca}^{2+}]_i$  in both  $\text{Ca}^{2+}$  release and uptake phases in muscle samples from human hearts [102]. Further, other research by

Connelly and colleagues, revealed substantial reduction of SERCA-2a ATPase in diabetic Ren-2 rats [109]. Upon examining the abundance of the inhibitory protein PLB, which is a regulator of SERCA 2a ATPase, a reduction of the active, phosphorylated form of PLN was observed in the diabetic state that is similar to that seen in the human hearts. Thus, it was predicted to reduce calcium transport and prevent actomyosin dissociation contributing to delayed relaxation and reduced contractility [110]. Lacombe et al. [111] reported that diabetes-induced diastolic dysfunction together with preserved overall systolic performance is coupled with abnormalities of intra-myocyte calcium regulation. Their findings included, prolonged  $\text{Ca}^{2+}$  transient decay, reduced intra-SR  $\text{Ca}^{2+}$  stores, reduced  $\text{Ca}^{2+}$  sparks, and decreased SERCA2a protein content, which were all consistent with decreased SR  $\text{Ca}^{2+}$  reuptake during the relaxation phase of cardiac myocytes. The decrease in SR  $\text{Ca}^{2+}$  load, combined with decreased ECC efficiency may contribute to the decrease in  $\text{Ca}^{2+}$  transient amplitude and  $\text{Ca}^{2+}$  spark frequency. One alternative mechanism for this approach could be defined as a compensatory mechanism *via* reduced SR  $\text{Ca}^{2+}$  leak which will enhance SR  $\text{Ca}^{2+}$  load [103,112] leading to the conclusion that impaired calcium reuptake during the diastolic phase, results from an impaired SERCA pump function.

Dhalla et al. [58] reported that activation of sympathetic nervous system and RAAS as well as platelet aggregation, endothelial dysfunction and generation of oxidative stress promote differential changes in SR activities and protein content during the development of diabetes. This results in alterations in SR function and SR remodeling occur in the diabetic heart. On the basis of these observations, it is suggested that oxidative stress and subcellular remodeling due to hormonal imbalance and metabolic defects play a critical role in the genesis of heart failure during the development of DCM. Remodeling of other subcellular organelles, including sarcolemma and myofibrils, has also been reported during the development of DCM [113-114].

### **Reduced calcium transients in diabetes mellitus**

During the last decade, accumulating evidence has been presented revealing that altered calcium homeostasis is of significant relevance for the pathophysiology of myocardial dysfunction and HF. Moreover, various mechanisms have been postulated as a result of many clinical and experimental studies. In addition, many outcomes have been identified from reduced  $\text{Ca}^{2+}$  transients and altered intracellular calcium cycling, reduced amplitude of  $\text{Ca}^{2+}$  transients with reduced systolic calcium concentrations and increased diastolic calcium

levels in isolated myocytes, together with decreased rates of SR calcium uptake in the failing human heart in type 1 DM [115-119]. Abnormal  $[Ca^{2+}]_i$  homeostasis has also been implicated in DCM and may precede clinical manifestation. Studies in cardiomyocytes have shown that diabetes results in impaired  $[Ca^{2+}]_i$  homeostasis due to altered SERCA and NCX activity [120]. Belke et al. [121] investigated contractile performance and  $Ca^{2+}$  transients in Langendorff-perfused hearts and isolated cardiac myocytes. They showed that in diabetic mouse hearts there was a decrease in rates of contraction, relaxation, and pressure development along with  $Ca^{2+}$  transients; significantly lower diastolic and systolic levels of calcium in myocytes from diabetic hearts. Furthermore, the decay rate of the  $Ca^{2+}$  transient was significantly reduced in diabetic myocytes, suggesting a diminished capacity for cytosolic calcium removal not associated with a change in NCX activity. Their study revealed that this decrease in contractile performance of the insulin-resistant (T2DM) diabetic model parallels the decrease in contractile performance observed in insulin-deficient (T1DM) diabetic models. However, recent study by Salem et al. [122] demonstrated no significant alterations in  $Ca^{2+}$  transients and L-type  $Ca^{2+}$  current in ventricular myocytes from 10-11 months old Goto Kakizaki (GK) sedentary compared to control sedentary rats or by exercise training (2-3 months of treadmill exercise). A study by Zhang et al. [123] found no evidence to support the idea that altered  $Ca^{2+}$  homeostasis underlies the contractile deficit of DCM. They postulated that the slower action potential and reduced SERCA2a expression could explain the slower  $Ca^{2+}$  transient kinetics in diabetic rats, but not the contractile deficit. Instead, they suggest that the observed LV remodelling may play a crucial role that is explained elsewhere [124-125].

In conclusion, the observed changes in contractility and in  $[Ca^{2+}]_i$  handling are most likely attributable to functional disturbances of SERCA2a, NCX and RyR2 in this transitional phase of diabetes.

## **Heart failure with normal ejection fraction**

Ejection fraction is the volume of blood (usually presented as a percentage) pumped out of the ventricles during each heartbeat or cardiac cycle. It is now extensively acknowledged that the clinical features of HF can occur in patients with normal left ventricular ejection fraction (LVEF) [126-128] a complex broadly referred to as HF with normal left ventricular ejection fraction (HFnEF). The finding of a reduced LVEF in patients with typical signs and

symptoms of HF provides objective proof that the patients suffer from cardiac abnormality and makes it obvious that the patient's clinical syndrome is indeed HF.

LV diastolic dysfunction is most commonly observed among patients with hypertension as well as the elderly [129]. While asymptomatic advanced LV diastolic dysfunction is also predictive of the future occurrence of HF [130], it is not clear why a number of patients with LV diastolic dysfunction have HFnEF while others remain asymptomatic. LV diastolic dysfunction has been suggested as the initial manifestation of diabetic heart disease in both T1DM and T2DM patients. In the Strong Heart Study [131], which investigated the effect of DM on LV filling pattern in normotensive and hypertensive individuals, T2DM was associated with an impaired relaxation pattern independently of age, blood pressure (BP), LV mass and LV systolic function.

The occurrence of DM in HF is approximately 20-35% [132] and is to some extent higher in patients with HFnEF, at 30-40% [133]. HFnEF and T2DM commonly coexist. Both conditions are associated with hypertension, obesity and ageing [23], all of which promote the prevalence of HF. However, irrespective of underlying CVD, DM has a central role in HFnEF as demonstrated in the CHARM study [134] whereby the relative risk of cardiovascular death or HF hospitalization conferred by DM was significantly greater in patients with HFnEF compared with those with HFrEF [134]. The prevalence of HFnEF may be high even in asymptomatic and well-controlled diabetic subjects [135,136]. Regardless of the increased prevalence and poor outcome, pathophysiological features underlying HFnEF in DM remain uncertain [137]. A recent study by Ehl et al. [138] demonstrated a significantly lower LVEF in diabetic compared with non-diabetic patients ( $P<0.001$ ) in a large patient population. Even though the difference was small, this finding may have important epidemiological impact, since LVEF is one of the most important predictors of survival. This hypothesis is supported by data from the Mayo clinic, which have confirmed a significantly worse survival of even asymptomatic diabetic patients with an LVEF of 50% [139].

Various morphological changes occur in the diabetic heart leading to the abnormal physiological findings described above. These include increased extracellular collagen deposition, interstitial fibrosis, myocyte hypertrophy and intra-myocardial micro-angiopathy. These changes are probably secondary to altered myocardial glucose and free fatty acid metabolism (FFA) in DM as outlined in Figure 3.



DM is also known to be associated with a loss of cardiac myofibrils [144], which are the main contractile components for muscle contraction. Severe loss of myofibrils can lead to decreased contraction and cardiac output that are frequently reported in diabetic patients. Experiments conducted by Pierce and Dhalla [145], demonstrated that the basal ATPase activity of myofibrils from diabetic hearts was significantly lower than the controls over 8 weeks. Their results also highlighted: (i) the basal and  $\text{Ca}^{2+}$  stimulated ATPase activities in diabetic rats demonstrated a greater sensitivity to KCl than control preparations, (ii) the myofibrillar basal ATPase, unlike  $\text{Ca}^{2+}$  stimulated ATPase, in diabetic animals exhibited a greater sensitivity to ethylene glycol and (iii) supports the view regarding the presence of some subtle structural and conformational changes in diabetic myofibrils. In subsequent work [146], diabetes was introduced in rats where they were maintained in a diabetic state for 6 weeks and then given 2 weeks of insulin treatment *in vivo*. The results obtained emphasized: (i)  $\text{Mg}^{2+}$ -ATPase and  $\text{Ca}^{2+}$ -stimulated ATPase activities were depressed in diabetic rat hearts in comparison to control, (ii) the depression in myofibrillar ATPases was of gradual onset as no changes were detected 2 weeks after inducing diabetes, (iii)  $\text{Mg}^{2+}$ -ATPase activity of myofibrillar preparations from control and diabetic hearts responded differently to N-ethylmaleimide modification, and interestingly and (iv) myofibrillar sulfhydryl reactivity to 5,5'-dithiobis (2-nitrobenzoic acid) was significantly depressed in diabetic preparations in comparison to control and insulin-treated diabetic animals.

While many studies focused on animal studies, Jweied and colleagues [147] conducted a study to determine whether human diabetes mellitus is associated with depressed cardiac myofilament function. Assessment of Myofilament function was achieved by determination of the developed force- $\text{Ca}^{2+}$  concentration relation in skinned cardiac cells from flash frozen human biopsies. Separate control experiments revealed that flash freezing of biopsy specimens did not affect myofilament function. In this study, all patients in the DM group were classified as T2DM patients, and most showed signs of diastolic dysfunction. Data from the results revealed that DM was associated with depressed myofilament function, that is, decreased  $\text{Ca}^{2+}$  sensitivity (29%,  $P < 0.05$  vs. control) and a trend toward reduction of maximum  $\text{Ca}^{2+}$ -saturated force (29%,  $P = 0.08$  vs. control). Results from this study highlighted that human diabetes mellitus is associated with decreased cardiac myofilament function and further, depressed cardiac myofilament  $\text{Ca}^{2+}$  responsiveness may underlie the decreased ventricular function characteristic of human diabetic cardiomyopathy.

## **Prevention, treatment and potential targets for DCM**

In order for proper and improved management options to be effective, it is essential to have better understanding of pathophysiology and pathogenesis in patients with DCM. These options include; lifestyle modification, glycaemic control, management of coexistent hypertension and heart failure. Some of aforementioned management options, in addition to emerging treatment modalities are discussed in the following section.

### **Lifestyle modification**

Exercise training is necessary to maintain a healthy body and it is recognized that regular exercise with better diabetic control would have beneficial effects on the disease outcome. In a recent study [148] physical activity was associated with a significant reduction in cardiovascular disease and all-cause mortality in patients with DM in many clinical studies. Exercise training was beneficial for reducing the incidence of DCM in both human patients and animal models [149,150]. Weight loss, control of fat and regular physical activity can positively adjust metabolic abnormalities and thus improve systemic insulin resistance. Insulin resistance can be achieved by increasing post-receptor-signaling and increasing insulin-mediated glucose transport, which seems to be associated with signal transduction at the level of phosphatidylinositol 3-kinase and insulin receptor substrate [151,152]. In addition to exercise, maintaining healthy eating patterns that are suitable for diabetic patients can also be expected to show similar beneficial effects.

### **Glycaemic control**

Clinical trials have demonstrated that poor glycaemic control has been associated with an increased risk of cardiovascular mortality – with an increase of 11% for every 1% rise in glycated haemoglobin (HbA<sub>1c</sub>) level [153]. It is expected that the effects of tight glycaemic control would be beneficial to patients because microvascular disease plays an important pathogenic role in the development of DCM. In one case-controlled study using cardiac MRI in patients with T1DM it was demonstrated that rigorous glycemic control was associated with better DCM outcome parameters [154] while in animal studies, improved glycaemic control delayed DCM in animal models [155]. However, other studies (UK Prospective Diabetes Study) failed to show a significant benefit of intensive blood glucose control using either sulphonylureas or insulin on the risk of developing macrovascular disease in patients with Type II diabetes [156].

## Anti-diabetic medication

There are various classes of anti-diabetic drugs, and their selection depends on the nature of the diabetes, age, as well as other factors. Some of the more popular and effective drugs will be discussed in this section. These include: metformin, thiazolidinediones, empagliflozin, GLP-1 and DPP-4.

Metformin, believed to be the most widely used medication for diabetes that is taken by mouth, upregulates cardiac autophagy, which is linked to DCM prevention and is primarily used to treat T2DM. Metformin facilitates glucose uptake and GLUT4 translocation in insulin-resistant cardiomyocytes and the myocardium by activating 5'-adenosine monophosphate-activated protein kinase [157]. This drug was reported to reduce mortality and improve the clinical outcome in overweight patients with HF and DM, however, metformin increase the production of lactate in the large intestine, which could potentially contribute to lactic acidosis [158].

Thiazolidinediones (TZDs) are a class of insulin sensitizers compounds for treating patients with T2DM. They act by increasing insulin sensitivity in skeletal muscle and adipose tissue through binding and activation of PPAR- $\gamma$ , a nuclear receptor that has a regulatory role in differentiation of cells [159].

Apart from insulin-sensitizing fat and skeletal muscle, TZDs increase the expression and function of glucose transporters in the heart, leading to improved glucose metabolism, and reduce non-esterified fatty acids (NEFA) utilization by the myocardium [160]. However, thiazolidinedione therapy can cause chronic symptoms that resemble heart failure. Therefore, the drug is generally not recommended in patients with heart failure.

Empagliflozin is a new anti-diabetic drug of the gliflozin class that was approved for treatment of T2DM in 2014. It is an inhibitor of sodium-glucose co-transporter 2 (SGLT-2), which reduces HbA<sub>1c</sub> levels in patients with T2DM by controlling visceral adiposity, blood pressure, arterial stiffness, albuminuria, weight, oxidative stress, hyperinsulinemia, and circulating uric acid levels [161].

Glucagon-like peptide-1 (GLP-1) is a known incretin that has the ability to decrease blood sugar levels in a glucose-dependent manner by enhancing the secretion of insulin. Synthetic



GLP-1, which has a longer half-life than natural GLP-1, has shown to attenuate myocardial apoptosis and enhance vasodilation [162]. This new group of drugs has shown promise in the management strategy in obese T2DM patients with DCM and further, GLP-1-based treatment has been associated with weight loss and lower hypoglycemia risks.

Dipeptidyl peptidase 4 (DPP-4) is an enzyme that metabolizes endogenous GLP-1. DPP-4 inhibitors can prevent cardiac diastolic dysfunction and cardiac hypertrophy by inhibiting fibrosis and oxidative stress in mouse models of insulin resistance and obesity [163].

## **Statins**

Statins (hydroxymethylglutaryl CoA reductase inhibitors) is a class of drug that lowers the level of cholesterol in the blood by reducing the production of cholesterol by the liver. Statins works by inhibiting HMG-CoA reductase (3-hydroxy-3-methyl-glutaryl-coenzyme A reductase), an enzyme found in liver tissue that plays a key role in the production of cholesterol in the body.

In multiple clinical trials [164] it was found that the use of statins reduces cardiovascular mortality and events in patients with diabetes and vascular risk factors, and it is beneficial even for primary prevention in patients without established cardiovascular disease [165].

It has been reported that atorvastatin, a lipid-lowering agent, reduces myocardial fibrosis, intramyocardial inflammation, and improves LV function in rat models of experimental DCM, independently of its LDL-C-lowering capacity [166]. Likewise, fluvastatin is useful for attenuating cardiac dysfunction and myocardial interstitial fibrosis in rat models of the disease [167].

## **Vasoactive medications - blockers and inhibitors**

Many vasoactive medications have been tried in both animal and human patients models with DCM, with variable results.

$\beta$ -blocker is a class of medications that are particularly used to manage abnormal heart rhythms, and to protect the heart from myocardial infarction after a first heart attack.

Initially, there was a reluctance to use  $\beta$ -blockers in patients with diabetes for fear of adverse effects on insulin resistance and an unawareness of hypoglycaemia. However, with the recent advances in the understanding of HF and the realization of the importance of the SNS in the release of vasoactive substances, they have become an essential treatment for HF [168].

Earlier studies of  $\beta$ -blockers in the early 90's involved recruited patients with advanced HF and, although LV function improved, mortality did not [169,170]. However, subsequent studies enrolled patients with mild-to-moderate HF and showed significant reductions in mortality rate of 32% and 34 % respectively [171,172]. The third generation  $\beta$ -blocker carvedilol, which antagonizes both  $\alpha$  and  $\beta$  receptors, has been proven to have a highly significant effect on mortality (67% reduction) in patients with severe HF [173]. Other  $\beta$ -blocker such as bisoprolol and metoprolol have been shown in large-scale randomized controlled trials to reduce heart failure mortality [174] and more recently, Sharma et al. demonstrate that  $\beta$ -Adrenoreceptor blockers were effective in experimental models of DCM [175].

The importance of (RAAS) antagonism in the prevention of diabetic CVD has demonstrated the key role that the RAAS plays in diabetic CVD onset and development. Angiotensin converting enzyme (ACEi) inhibitors and angiotensin II receptor blockers (ARB) represent the first line therapy for primary and secondary CVD prevention in patients with diabetes [176]. (ACEi), renin inhibitor (aliskiren) and (ARB) were all shown to be protective against DCM in rat models [177]. Other studies have demonstrated that ACEis and ARBs were also beneficial in both human and animal models of DCM [178,179].

Recently, Poly (ADP-ribose) polymerase (PARP) inhibitors have stimulated much excitement. PARP-1 is a member of the PARP enzyme family and is one of the most abundant nuclear proteins which functions as a DNA-nick-sensor enzyme [180]. Research conducted by Du et al. [181] revealed that hyperglycemia-induced overproduction of superoxide by the mitochondrial electron transport chain activates the three major pathways of hyperglycemic damage found in aortic endothelial cells by inhibiting GAPDH (glyceraldehyde-3-phosphate dehydrogenase) activity. Thus, inhibition of PARP blocks hyperglycemia-induced activation of multiple pathways of vascular damage and provides a unique approach as it blocks activation of all the major pathways thought to mediate tissue damage in diabetes.

### **Metabolic modulators**

Metabolic modulators are a newer class of drugs that benefit patients by modulating cardiac metabolism without altering hemodynamics. They have the potential to relieve symptoms in patients with refractory heart failure who are already on optimal medical therapy. These drugs increase glucose metabolism at the expense of free fatty acid metabolism, thereby enhancing efficient use of oxygen. Three metabolic modulators drugs that could potentially

be used for HF therapy and CDM are: trimetazidine, resveratrol and ranolazine.

Trimetazidine, a competitive inhibitor of the terminal enzyme in  $\beta$ -oxidation has promising beneficial effects on heart failure in diabetic patients with both idiopathic and ischemic dilated cardiomyopathy [182]. Studies of animal models revealed that trimetazidine reduce free radical injury, improve endothelial function, inhibit apoptosis and attenuate lipotoxicity [183]. However, human trials are needed to investigate the beneficial effects of this drug on the treatment and prevention of DCM. Studies on resveratrol revealed that it reduces glucose levels; improve triglyceride level, heart rate, and glycemia [184] while ranolazine, a potent late  $\text{Na}^+$  current inhibitor, normalize altered cardiomyocyte intracellular calcium concentration due to the close relationship between  $\text{Ca}^{2+}$  and  $\text{Na}^+$  handling by the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger [186].

### **New therapeutic directions**

Within the past five years, research into new therapeutic direction focused on the areas of cell/genetic therapy and targeting mitochondrial oxidative stress. A few of these new emerging treatment modalities will be discussed in this section. An excellent review on this topic was done by Huynk et al. [186] in which they discussed both conventional and novel therapeutic approaches for the treatment of left ventricular dysfunction in diabetic patients.

mi-RNA.

The dysregulation of micro-ribonucleic acid (mi-RNA) function serve as an important pathogenic mechanism of diabetes and its complications. mi-RNAs have been reported to be potential biomarker for early detection of DCM and they have become the subject of an active area of research to establish their potential contribution to heart disease in patients with T2DM. The noncoding RNA, miR-223 was found to be associated with regulation of GLUT4 expression in adult cardiomyocytes [187] while data from experimental studies underscore the contribution of miR-21 in stimulating MAP kinase signaling in mouse fibroblasts, consequently promoting fibrosis and contractility alterations as features of DCM in diabetic animal models [188,189]. Other research have implicated miRNA to be involved in regulating extracellular signal regulated kinases (ERK) in diabetic conditions and therefore, modulating ERK1/2 derived-pathway which opposes oxidative stress-induced insulin resistance in cardiomyocytes [190].

#### Coenzyme Q10.

Recent research by Mortensen et al. [191], on the effect of coenzyme Q10 on morbidity and mortality in chronic heart failure patients using randomized controlled trial, showed that long-term coenzyme Q10 treatment in patients with chronic heart failure is safe, improves symptoms, and reduces all-cause mortality by 42% and cardiovascular death by 43%. Other reports have also established that coenzyme Q10 improves cardiac function in patients with DM and concurrent heart failure [192,193].

#### Pim-1.

Pim-1 (serine/threonine-protein kinase pim-1) gene therapy was shown to improve LV diastolic function, prevent cardiac apoptosis, fibrosis, and development of HF [194]. While in other studies [195], it was demonstrated that Pim-1 downregulation contributes in the pathogenesis of diabetic cardiomyopathy. Thus, establishing that intravenous gene therapy with pim-1 *via* a cardiotropic viral vector halts the progression of diabetic cardiomyopathy through promotion of pro-survival signaling and represents a novel and effective approach to treat the disease.

#### Phosphoinositide 3-kinase.

In a very recent research, Prakoso and colleagues [196] investigated the therapeutic potential of a delayed intervention with cardiac-targeted phosphoinositide 3-kinase (PI3K) gene therapy, administered to mice with established diabetes-induced LV diastolic dysfunction. After study endpoint, it was discovered that diabetes-induced LV dysfunction was significantly attenuated by a single administration of recombinant adeno-associated-virus 6-constitutively active PI3K (p110 $\alpha$ ) (rAAV6-caPI3K), administered 8 weeks after the induction of diabetes. Their results clearly demonstrate that cardiac-targeted PI3K (p110 $\alpha$ ) gene therapy limits diabetes-induced up-regulation of NADPH oxidase and cardiac remodelling suggests new insights into promising approaches for the treatment of diabetic cardiomyopathy.

#### Szeto-Schiller peptide.

The SS (Szeto-Schiller) peptide antioxidants represent a novel approach with targeted delivery of antioxidants to the inner mitochondrial membrane. The structural motif of these SS peptides centers on alternating aromatic residues and basic amino acids (aromatic-cationic peptides). These SS peptides can scavenge hydrogen peroxide and peroxynitrite and

inhibit lipid peroxidation. By reducing mitochondrial ROS, these peptides inhibit mitochondrial permeability transition and cytochrome c release, thus preventing oxidant-induced cell death [197].

The Szeto-Schiller peptide d-Arg-2', 6'-dimethyltyrosine-Lys-Phe-NH<sub>2</sub> (SS31) is a positively charged free-radical scavenger that can accumulate to high levels in the mitochondria and prevent diastolic dysfunction, fibrosis, and cardiac hypertrophy [192].

This technique represents a promising approach for preventing DCM by targeting excess myocardial ROS with novel antibiotics.

## **Conclusion**

The prevalent rise in DM worldwide has made DCM an increasing health concern. As the incidence and prevalence of DM continue to rise, HG-induced structural and ultrastructural changes may cause a reduction in heart perfusion and eventually HF. A variety of treatment options have shown to be effective in treating DCM and novel therapeutic strategies, such as gene therapy targeting the phosphoinositide 3-kinase PI3K (p110 $\alpha$ ), signaling pathway, and miRNA dysregulation have shown good promise. In addition to these, targeting redox stress and mimetic peptides targeting calcium channels may represent a future strategy for combating the ever-increasing incidence of heart failure in the diabetic population.

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